recrystallization from Et₂O gave 12 (1.66 g, 97%) as colorless needles: mp 93–94 °C (Et₂O); IR (CH₂Cl₂) 3600, 2920, 2870, 1720 cm⁻¹; ¹H NMR (CDCl₃) δ 0.76 (d, 3 H, J = 1.0 Hz, 4a-CH₃), 1.36–2.60 (m, 13 H), 3.48 (dd, 1 H, $J_{4,3} = 11.2$ Hz, $J_{4,3'} = 4.5$ Hz, 4-H), 3.70 (s, 3 H, CO₂CH₃); MS, M⁺ 240.1364, calcd for C₁₃H₂₀O₄ 240.1361. Anal. Calcd for C₁₃H₂₀O₄: C, 64.98; H, 8.39. Found: C, 64.79; H, 8.38.

Methyl $(1\beta, 4\beta, 4a\beta, 8a\alpha)$ -4a-Methyl-6-oxo-4-[(p-tolylsulfonyl)oxy]-1,2,3,4,4a,5,6,7,8,8a-decahydro-1-naphthoate (3). To a stirred solution of 12 (500 mg, 2.08 mmol) in dry CH₂Cl₂ (15 mL) at 0 °C under N_2 was added anhydrous pyridine (0.33 mL, 4.1 mmol) followed by p-toluenesulfinyl chloride²³ (0.38 mL, 4.1 mmol). The mixture was stirred for 3 h at 0 °C, then poured into 1 N HCl, and extracted with CH₂Cl₂. The combined extracts were washed with 5% NaHCO3 solution, water, and brine and dried over $MgSO_4$. Evaporation of the solvent in vacuo gave a mixture of diastereomeric sulfinate esters (786 mg) as a colorless oil: IR (CH₂Cl₂) 2870, 1705, 1130, 1115, 840 cm⁻¹; ¹H NMR (CDCl₃) δ 0.76 and 0.78 (2 s, 3 H, 4a-CH₃), 1.28-2.40 (m, 11 H), 2.42 (s, 3 H, ArCH₃), 2.54-2.68 (m, 1 H, 1-H), 3.67 and 3.70 (2 s, 3 H, CO₂CH₃), 4.09 and 4.20 (2 dd, 1 H, J = 4.40, 12.2 Hz, 4-H), 7.38 and 7.65 (2 d, 2 H each, J = 8.0 Hz, Ar). To a stirred solution of the crude sulfinate ester (786 mg, 2 mmol) in dry CH₂Cl₂ (15 mL) at 0 °C under N₂ was added *m*-chloroperbenzoic acid (500 mg, 2.9 mmol). After having been stirred for 3 h at 0 °C, the mixture was poured into saturated NaHCO3 solution, washed with water and brine, and dried over MgSO4. Evaporation of the solvent in vacuo and purification of the residue by flash chromatography (PhH-Et₂O-petroleum ether, 1:2:2) gave the tosylate 3 (656 mg, 80%) as colorless crystals: mp 153–154 °C (Et₂O); IR (CH₂Cl₂) 2850, 1725, 1355, 1180, 840 cm⁻¹; ¹H NMR (CDCl₃) δ 0.78 (s, 3 H, 4a-CH₃), 1.54-2.40 (m, 11 H), 2.46 (s, 3 H, ArCH₃), 2.54-2.64 (m, Hz, H, 1-H), 3.68 (s, 3 H, CO₂CH₃), 4.45 (dd, 1 H, J = 4.40, 12.18 Hz, 4-H), 7.40 and 7.84 (two d, 2 H each, Ar); MS, m/e (relative intensity) 394 (7, M⁺), 239 (12), 223 (base), 222 (30), 207 (44). Anal. Calcd for $C_{20}H_{26}O_6S$: C, 60.89; H, 6.64; S, 8.13. Found: C, 60.70; H, 6.61; S, 8.05.

Methyl $(1\beta,4\beta,4a\beta,8a\alpha)$ -4a-Methyl-6-oxo-7-[2-hydroxy-3-[(tetrahydropyranyl)oxy]propanyl]-1,2,3,4,4a,5,6,7,8,8adecahydro-1-naphthoate (13). To a stirred solution of LDA (0.97 mmol) in dry THF (2 mL) at -78 °C under N₂ was added dropwise a solution of 3 (175 mg, 0.44 mmol) in dry THF (4 mL). After 0.5 h, anhydrous zinc chloride (137 mg, 1.01 mmol) in dry THF (2 mL) was added. After another 5 min, 1-[(tetrahydropyranyl)oxy]-2-propanone⁷ (153 mg, 0.97 mmol) in dry THF (1 mL) was added, and the stirring was continued at -78 °C for 0.5 h and then at -20 °C for 1 h. Aqueous NH₄Cl solution was added, and the product was extracted with Et₂O. The combined extracts were washed with water and brine and dried over MgSO₄. Evaporation of the solvent in vacuo followed by flash chromatography (PhH-Et₂O-CH₂Cl₂, 2:2:1) of the residue gave unreacted 3 (17 mg, 10%), the THP adduct 13 (121 mg, 52%), and the

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dialkylated adduct 14 (109 mg, 35%), both as colorless oils. THP adduct 13: IR (CH₂Cl₂) 3500, 2940, 2870, 1705, 1695, 1360, 1180, 840, 820 cm⁻¹; ¹H NMR (CDCl₃) δ 0.73, 0.76, 0.86, 0.89 (4 s, CH₃), 1.22 and 1.27 (2 s, R'(CH₃)CHOHR"), 1.56–2.30 (m, 18 H), 2.47 and 2.48 (2 s, ArCH₃), 3.34–3.94 (s overlapping m, 7 H, CO₂CH₃ and THP), 4.41 (dd, 1 H, J = 4.38, 12.20 Hz, 4-H), 4.54–4.72 (m, 2 H, R'(Me)CHOHR" and THP), 7.38 and 7.82 (2 d, 2 H each, J = 8.0 Hz, Ar); MS, m/e (relative intensity) 506 (0.1, M – CH₃), 190 (3), 172 (62), 155 (7), 108 (25), 107 (33), 91 (base). Dialkylated adduct 14: IR (CH₂Cl₂) 3500, 2930, 2860, 1725, 1360, 1180, 840, 820 cm⁻¹; ¹H NMR (CDCl₃) δ 0.82–0.96 (m, CH₃), 1.08–1.40 (m, 6 H, two CH₃), 1.44–2.40 (m, 24 H), 2.47 and 2.56 (2 s, ArCH₃), 3.30–3.93 (s overlapping m, 11 H, CO₂CH₃ and THP), 4.40–4.72 (m, 5 H, R'(Me)CHOHR", THP, and 4-H), 7.34–7.50 (m, 2 H, Ar), 7.70–7.94 (m, 2 H, Ar).

 $(4a\alpha,5\beta,8\beta,8a\beta)$ -5-(Methoxycarbonyl)-3,8a-dimethyl-8-[(ptolylsulfonyl)oxy]-4,4a,5,6,7,8,8a,9-octahydronaphtho[2,3b]furan (2). A stirred solution of 13 (40 mg, 0.08 mmol) in THF-H₂O (2:1, 1.5 mL) containing a catalytic amount of ptoluenesulfonic acid (1 mg) was heated at 60 °C for 0.5 h. The solution was cooled and poured into aqueous NaHCO₃ solution, and the product was extracted with Et_2O . The combined ethereal extracts were washed with water and brine and dried over MgSO4. Evaporation of the solvent in vacuo, followed by immediate flash chromatography (CH_2Cl_2) , gave the furan 2 (6 mg, 18%) as a colorless oil which is unstable and decomposes slowly on standing at room temperature: IR (CHCl₃) 2933, 2845, 1729, 1597, 1175, 1019, 940 cm⁻¹; ¹H NMR (CDCl₃) δ 0.87 (s, 3 H, 8a-CH₃), 1.91 (d, $3 H, J = 1.16 Hz, C3-CH_3), 1.55-2.66 (m, 10 H), 2.46 (s, 3 H)$ $ArCH_3$, 3.69 (s, 3 H, CO_2CH_3), 4.44 (dd, 1 H, J = 4.41, 12.16 Hz, 8-H), 7.01 (br s, 1 H, 2-H), 7.40 and 7.86 (2 d, 2 H each, J = 8.0Hz, Ar); MS, m/e (relative intensity) 432 (11, M⁺), 261 (2), 260 (6), 155 (31), 108 (57), 91 (base).

Sericenine (1). To the furan 2 (12 mg, 0.03 mmol) in dry THF (1 mL) was added potassium bis(trimethylsilyl)amide in THF (0.06 mL, 0.06 mmol). After stirring for 1 h at 25 °C under N₂, the reaction had gone to completion by TLC. The mixture was poured into water and extracted with Et_2O . The ethereal extracts were washed with water and brine and dried over MgSO₄. Evaporation of the solvent in vacuo followed by flash chromatography $(CH_2Cl_2$ -petroleum ether, 3:7) of the residue gave 1 as white needles (3 mg, 43%): mp 111–114 °C (lit.⁴ mp 115 °C); IR (CHCl₃) 3040, 1706, 1640, 1550 cm⁻¹; ¹H NMR (CDCl₃) δ 1.36 (br s, 3 H, 14-CH₃), 1.97 (d, 3 H, J = 1.16 Hz, 13-CH₃), 2.2–2.64 (m, 4 H), 2.78-2.94 (m, 1 H, 7- β H), 3.15 and 3.54 (2 d, 1 H each, J = 14.7Hz, 10β , 10α -H), 3.52-3.66 (br d, 1 H, 7α -H), 3.73 (s, 3 H, CO₂CH₃), 5.50 (t, 1 H, J = 7.5 Hz, 2-H), 6.63 (br d, 1 H, 6-H), 7.06 (br s, 1 H, 12-H); MS, m/e (relative intensity) 260 (45, M^+), 245 (11), 229 (6), 201 (19), 108 (base), m/e calcd for $C_{16}H_{20}O_3$ 260.1412, found 260.1422. Comparison of the IR, ¹H NMR, and MS with the corresponding spectra of natural sericenine⁴ was identical.

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Ultrasound in Phosphine Preparation. Simple Preparations of Some Bis(alkylphenylphosphino)alkanes

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Diphosphines of the class RPhP(CH₂)_nPPhR (n = 2-6) have been prepared from the reactions of the corresponding Ph₂P(CH₂)_nPPh₂ with alkali metal followed by alkylation. Ultrasound irradiation should be applied in the reductive cleavage stage so as to assure the purity of the final products.

Diphosphines of the general type $PhRP(CH_2)_nPRPh$ are important chelating reagents for the preparation and mechanistic study of various transition-metal complexes and catalysts. The conventional method preparation in-

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volves the deprotonation and subsequent alkylation of a secondary diphosphine HPhP(CH₂), PPhH, prepared from PhP²⁻ or HPhP⁻ and a α, ω -dihaloalkane.² These preparations are expensive and multistep and require the manipulation of extremely air-sensitive intermediates such as secondary phosphines. Obviously, simpler approaches toward the preparation of this class of compounds are desired.

It is well established that the P-Ph bond of triphenylphosphine³ and an alkyldiphenylphosphine⁴ can be cleaved by an alkali metal to the corresponding phenyl phosphide anion and then can be alkylated with electrophiles to give the substituted tertiary phosphines. However, the double reductive cleavage of the P-Ph bonds at the two ends of a bis(diphenylphosphino)alkane (diphos) has not been achieved although this appears the most convenient source of "PhP(CH₂)_nPPh", the requisite precursor to the diphosphine $RPhP(CH_2)_nPPhR$. We therefore investigated the possibility of the double reductive cleavage of the P-Ph bond of diphos 1 (n = 2-6) with alkali metals.

The reaction of a slight excess of K with the diphos 1a (n = 2) was completed in 20 min at 0 °C, giving a dark red solution, indicating the formation of certain type of phosphide anion. After destroying the phenyl anion with t-BuCl, 2 equiv of MeI were added, yielding a complex mixture in which 2a was the major component (eq 1). By

$$Ph_{2}P(CH_{2})_{n}PPh_{2} \xrightarrow{i} [^{Ph}P(CH_{2})_{n}PPh^{-}] + [Ph^{-}] \xrightarrow{Hi,Hi}$$

$$1a, n = 2$$

$$b, n = 3$$

$$c, n = 4$$

$$d, n = 5$$

$$e, n = 6$$

$$PhMeP(CH_{2})_{n}PMePh + MePPh_{2} + impurities$$

$$2a, n = 2$$

$$3$$

(1)

(i) K, O °C; (ii) /-BuCl; (iii) MeI.

TLC and temperature-programmed mass spectral analyses, $MePPh_2$ (3) was found as a side product. The product mixture could not be purified by chromatography or fractional distillation. Lowering the reaction temperature down to -78 °C resulted in the product being less contaminated, but the reaction proceeded at a much slower rate. For example, 100 mg of 2a required 12 h for complete reaction. Similar difficulties were encountered for the reaction of K with 1b-e. Therefore, the attempt of using K for the double reductive cleavage was abandoned.

The use of Li instead of K with 1a-e in this reaction sequence at 0 °C gave similar, but slightly cleaner, products 2a-e at a very slow rate. Attempts to accelerate the reaction rate by using a large excess of Li or by raising the reaction temperature all caused the formation of the side product 3 and other unseparable impurities. If the reaction was carried out at higher temperature or by prolonged stirring, the ether linkage of the solvent THF was cleaved by the nucleophilic attack of the phosphide anion⁵ causing additional side products.

Since the reductive cleavage of the P-Ph bond of triphenylphosphine by metal Li can be accelerated by ultrasound irradiation.⁶ it was anticipated that a similar rate

enhancement should be observed on the reactions of diphosphines. Indeed, when a diphos 1a-e and a slight excess of Li were irradiated with ultrasound at 0 °C,7 the reaction took place immediately and was complete within minutes. After the addition of 2 equiv of t-BuCl, the alkyl halide (MeI or bromocyclohexane) was added to yield the disubstituted product 2a-e or 4a-e (eq 2). The result was

$$1a-e \xrightarrow{i, ii} (C_{6}H_{11})PhP(CH_{2})_{n}PPh(C_{6}H_{11})$$

$$4a-e(a+2-6)$$
(2)

20-0

excellent not only because the rate was enhanced but also because the product contained only one component. Further purification by rapid column chromatography or simple distillation gave the product in high yield (Table I). It was noted that once the addition of t-BuCl was completed, the alkyl halide should be added immediately. Otherwise, a major side product containing the P-t-Bu bond⁸ would form in significant quantity.⁸ The later the alkyl halide was added, the more this side product formed. Also, if alkyl halide was added without the addition of t-BuCl, the product mixture was even more complex. This agreed with the report that the phenyl anion would induce side reactions.⁶

The reductive cleavage reaction and alkylation reactions were successful for the diphosphines 1a-e. However, for bis(diphenylphosphino)methane (dppm) (5), the reductive cleavage proceeded via a completely different route. The reaction of 5 with either K or Li by usual stirring or ultrasound irradiation all failed to give the diphosphide dianion. Instead, the CH₂-P bond was cleaved as evidenced by the fact that $MePPh_2$ (3) was produced when the reaction was worked up by MeI (eq 3).



(i) Li, ultrasound irradiation; (ii) t-BuCl; (iii) MeI.

Tests were also made to control the amount of Li or K to about 2 equiv during the reduction with the diphosphines la-e in the expectation that only one P-Ph bond would be cleaved to yield the species Ph₂P- $(CH_2)_n PPh^-$ and, after alkylation, the unsymmetrical diphosphine $Ph_2P(CH_2)_nPRPh$ (6). However, after the Li was completely consumed and MeI added, only the disubstituted product 2a-e and the unreacted starting material la-e were detected in about a 1:1 ratio, while the desired monosubstituted product was barely detected by temperature-programmed mass spectral analysis. The inability to produce the monophosphide anion by this method might have been attributed to the heterogeneous

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⁽⁷⁾ An ordinary laboratory ultrasound cleaner was used. (Bransonic 220, Branson Cleaning Equipment Co. Shelton, CT).

⁽⁸⁾ We were unable to isolate this type of side products in pure states. However, in each case for 1a-e, the ¹H NMR spectrum showed a doublet around $\delta 0.95$ (J = 12 Hz), indicating that the existence of P-C(CH₃)₃ was highly possible. The mass spectrum showed a distinct peak at the mass number of PhHP(CH₂)_nPPhBu. The side products were thus speculated to be PhHP(CH₂)_nPPh(t-Bu), which are reasonable side products of the reactions of the diphosphine dianions with isobutene which came from t-BuCl.

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		physical data	bp _{1.0} 180-182 °C (lit. bp 250-260 °C ^a)	$p_{1,0} = 195-200 \ ^{\circ}C$ (lit. bp _{0.4} = 199-201 $\ ^{\circ}C^{b}$)	mp 42 °C, disulfide mp 135–139 °C (lit. disulfide mp 138–141 °C ^{b})	bp _{1,0} 255–262 °C (lit. bp _{0,6} 257–258 °C ^b)	mp 47–48 °C, disulfide mp 135–140 °C (lit. mp 45 °C, disulfide mp 130–150 °C ^b)	disulfide mp 252-256 °C	disulfide mp 198-203 °C	mp 102–104 °C. disulfide mp 242–245 °C (lit. mp 104–105 °C, disulfide mp 248–250 °C ^c)	disulfide mp 182-185 °C (lit. disulfide mp 184-185 °C ^c)	disulfide mp 210–214 °C (lit. disulfide mp 211–214 °C ^c)	(.; Krech, F. Chem. Ber. 1961, 94, 2656.
Table I. Preparation of Bis(alkylphenylphosphino)alkane	elemental analyses	found	C, 70.1; H, 7.4	C, 71.0; H, 7.5	C, 71.8; H, 7.9	C, 72.2; H, 8.2	C, 72.9; H, 8.4	C, 76.0; H, 8.6	C, 76.1; H, 8.9	C, 76.8; H, 8.9	C, 76.8; H, 9.3	C, 77.1; H, 9.5	^c Issleib, h
		calcd	C, 70.1; H, 7.3	C, 70.8; H, 7.6	C, 71.5; H, 7.9	C, 72.1; H, 8.2	C, 72.7; H, 8.5	C, 76.1; H, 8.8	C, 76.4; H, 9.0	C, 76.7; H, 9.1	C, 77.0; H, 9.3	C, 77.2; H, 9.4	96, 2186.
		MS, m/e	274 (M ⁺), 259 (base), 246, 227, 123, 121, 91	288 (M ⁺), 273 (base), 165, 123, 121, 109, 91	302 (M ⁺), 287 (base), 179, 123, 121, 109, 91	316 (M ⁺), 301 (base), 193, 123, 109, 91	330 (M ⁺), 315 (base), 208, 207, 193, 138, 123, 121, 109, 91	410 (M ⁺), 327 (base), 219, 217, 191, 109, 108	424 (M ⁺), 341 (base), 233, 191, 109, 108	438 (M ⁺), 355 (base), 247, 191, 178, 109, 108	452 (M ⁺), 369 (base), 261, 191, 178, 109, 108	466 (M ⁺), 383 (base), 275, 191, 109, 108	Gruber, K. <i>Chem. Ber.</i> 1963,
		IR, cm ⁻¹	3080, 2950, 1840, 1500, 1060	3080, 2950, 1840, 1500, 1060	3080, 2950, 1500, 1840, 1500, 1060	3080, 2950, 1840, 1500, 1060	3080, 2950, 1840, 1500, 1060	3080, 2950, 1500, 1840, 1500, 1060	3080, 2950, 1500, 1840, 1500, 1060	3080, 2950, 1500, 1840, 1500, 1060	3080, 2950, 1500, 1840, 1500, 1060	$\begin{array}{c} 3080,\ 2950,\\ 1840,\ 1500,\\ 1060\end{array}$	b, K.; Krech, F.; (
		NMR, δ	1.25 (d, 6 H, <i>J</i> = 2 Hz), 1.6-1.8 (m, 4 H), 7.5 (br s. 10 H)	1.25 (d, 6 H, J = 2 Hz), 1.4-1.75 (m, 6 H), 7.5 (br s, 10 H)	1.3 (d, 6 H, J = 2 Hz), 1.35-1.65 (m, 8 H), 7.5 (br s, 10 H)	1.25 (d, 6 H, <i>J</i> = 2 Hz), 1.25-1.75 (m, 10 H), 7.5 (br s, 10 H)	1.25 (d, 6 H, J = 2 Hz), 1.0-1.7 (m, 12 H), 7.5 (br s, 10 H)	1.0–2.4 (m, 26 H), 7.5 (br s, 10 H)	1.0-2.2 (m, 28 H), 7.5 (br s, 10 H)	1.0–2.0 (m, 30 H), 7.5 (br s, 10 H)	0.9-1.9 (m, 32 H), 75. (br s, 10 H)	0.9–1.8 (m, 34 H), 7.5 (br s, 10 H)	2r. 1963, 96, 279. ^b Issleib.
	yield, %		84	86	16	16	94	80	85	88	06	87	vem. Be
		prod.	2a	2b	2c	2d	2e	4a	4b	4c	4d	4e	K. Ch
	time. time		1				1	63	5	5	61	5	udtke
	alkvl	halide	MeI	MeI	MeI	MeI	MeI	C ₆ H ₁₁ Br	C ₆ H ₁₁ Br	C ₆ H ₁₁ Br	C ₆ H ₁₁ Br	C ₆ H ₁₁ Br	ib, K.; Sta
		diphos	1a	1b	1c	1d	le	1a	1b	1c	1d	le	^a Issle

nature of this type of reactions. Because of the heterogeneity, once the monophosphide anion was formed, it became the species that was closest to the Li metal. Therefore, the second P-Ph bond on the same molecule would be cleaved faster than that of another molecule. Therefore, the diphosphines 1a-e were either doubly cleaved or unreacted (eq 4).

 $1a-e \xrightarrow{i \text{ or } ii} \begin{cases} Ph_2 P(CH_2)_{/2} PPh^{-1} \\ 6 \\ iii, iv \end{cases}$ (4)

2a-e + recovered 1a-e

(i) 2 equiv of Li, ultrasound irradiation; (ii) 2 equiv of K, -78 °C; (iii) \neq -BuCl; (iv) MeI.

In conclusion, the double reductive cleavage of the P-Ph bond of a diphosphine can normally be achieved under mild conditions such as with K at -78 °C or with the less reactive Li at 0 °C at extremely slow rate. Subsequent alkylation gives the disubstituted diphosphine with contamination. Attempted rate enhancement by increasing the reaction temperature is unsatisfactory while ultrasonic irradiation not only accelerates the reaction but also eliminates the formation of side products. The reason why ultrasonic irradiation selectively increases only the rate of the main reaction and not that of the side reactions is still unclear. It is suspected, however, that the main product is derived from a heterogeneous reaction on whose rate the ultrasound irradiation normally has significant influence¹⁰ and that the side reactions may be derived from some type of homogeneous reactions on whose rate the ultrasound has little or no influence.

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Experimental Section

All reactions and workup procedures were manipulated under anhydrous nitrogen atmosphere. All solvents were dried and deoxygenated before use. NMR spectra were recorded on a JEOL C-60-HL NMR spectrophotometer. IR spectra were recorded on a Perkin-Elmer 297 infrared spectrophotometer. MS spectra were taken on a HP 5990B gas chromatograph/mass spectrometer. Elemental analyses were taken at National Taiwan University.

Preparations of the Diphosphines 2a-e and 4a-e. Since the procedures are similar, 2b is used here as a typical example. To a vial containing finely chopped metal Li (50 mg, 7.2 mmol) in THF (3 mL) cooled at -10 °C was added dropwise bis(diphenylphosphino)propane (dppp) (1b) (300 mg, 1.2 mmol) in THF (2 mL). The colorless reaction mixture was put into an ultrasound cleaner kept at 0 °C and irradiated. The reaction was followed by TLC (alumina, hexane/EtOAc) until the starting material was consumed (less than 10 min). The dark red solution was separated from the excess of Li metal, and t-BuCl (220 mg, 2.4 mmol) in THF (1 mL) was added immediately at room temperature. No color change was observed at this stage. The resulting solution was again cooled to 0 °C, and as MeI (340 mg, 2.4 mmol) was added, the color faded instantaneously. After the mixture was stirred for another 60 min, water (8 mL) was added, and the lavers were separated. The aqueous layer was extracted with benzene $(4 \times 10 \text{ mL})$. The combined organic layers were dried (Na₂CO₃), concentrated under reduced pressure and purified by flash column chromatography (alumina, benzene), giving the diphosphine 2b.

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Registry No. 1a, 1663-45-2; 1b, 6737-42-4; 1c, 7688-25-7; 1d, 27721-02-4; 1e, 19845-69-3; 2a, 23808-01-7; 2b, 98170-68-4; 2c, 98170-69-5; 2d, 98170-70-8; 2e, 98170-71-9; 4a, 98170-72-0; 4b, 98170-73-1; 4c, 72144-83-3; 4d, 90116-42-0; 4e, 98170-74-2; MeI, 74-88-4; $C_6H_{11}Br$, 108-85-0.

Conversion of Thiosulfinate Derivatives of Cystine to Unsymmetrical Cystines and Lanthionines by Reaction with Tris(dialkylamino)phosphines

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Synthetic routes to the unsymmetrical lanthionines 2 and 3 have been developed. Reaction of a thiosulfinate, derived by oxidation of the corresponding symmetrical cystine with m-chloroperbenzoic acid, with a protected cysteine in the presence of a tris(dialkylamino)phosphine, hexaethylphosphorus triamide, yielded an unsymmetrical or mixed cystine. Contraction of the disulfide linkage in the appropriate mixed cystine by reaction with hexaethylphosphorus triamide provided the unsymmetrical lanthionines 2 and 3. A second route to unsymmetrical lanthionines was investigated that involved the attempted displacement of sulfonate leaving groups from dior tripeptides containing a serylvaline residue. The sulfonate ester function was observed to be very unreactive toward displacement by mercaptide anion in these peptides, a fact that may be due to steric or conformational effects originating from the adjacent valine residue.

The quinomycin antibiotics¹ are a structurally unique group of cyclic depsipeptides that contain a novel β -methylthiolanthionine as a constituent of the peptide.

Echinomycin (1), the most prominent of the quinomycins, is known to bind to deoxyribonucleic acids by bifunctional intercalation and to thereby function as a potent inhibitor of RNA synthesis.² The antibiotics are known to possess

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